ANA BIO ISP CK - MB

(Immunoinhibition Method)

For Miura Instruments

Intended Use

CK- MB is a reagent set for determination of CK-MB activity in serum and plasma based on Immunoinhibition method.

Principle

The serum sample is incubated with CK-MB reagent containing antibody specific to CK-M subunit which completely inhibits the CK-M monomer. The activity of CK-B which is not inhibited by the antibody is then measured by the following reaction sequence.

Creatine phosphate + ADP Hexokinase ATP + Glucose glucose-6-phosphate + ADP

G-6-PDH

G-6-phosphate + NADP⁺ $\leftarrow \bullet$ 6-phosphogluconate + NADPH + H⁺

Components & Concentration of Re	eagents
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Reagent	Component	Concentration	
Reagent 1	Imidozole buffer pH 6.6	125 mmol/L	
	НК	≥ 800 U/L	
	Nac	25 mmol/L	
	G6P-DH	≥ 800 U/L	
	NADP	2.5 mmol/L	
	CK-MM Ab	≥ 1000 U/L	
	Mg Acetate	10 mmol/L	
	Stabilizers, excipients & surface active agents		
Reagent 2	СР	30 mmol/L	
	ADP	2 mmol/L	
	AMP	6 mmol/L	
	DAPP	0.010 mmol/L	
	Stabilizers, excipients & surface active agents		

Reagent storage and stability

The reagent kit should be stored at 2° - 8° C and is stable till the expiry date indicated on the label.

A slight variation in the composition of the components may occur between batches, but this has no effect on the test results. After opening, the vial R1 and R2 are stable 30 days if recapped immediately and protect from contamination, evaporation, direct light and stored at correct temperature.

Specimen collection and preservation

Blood should be collected in a clean dry container. Although serum is preferred, plasma with heparin or EDTA can be used. The assay should be carried out as far as possible on the same day. Serum and plasma samples are stable for 1 week at 4° C and 1 month at -10 °C. The samples should be brought to room temperature prior to use. Avoid use of haemolysed and grossly contaminated samples.

Automation

This kit, though developed and manufactured to be used as manual assay and with I.S.E. Miura Analyzer, can be used also with other analyzers able to meet the specifications indicated in section "Reaction conditions – Test procedure" Application sheets are available for automatic instruments.

All applications not explicitly approved by KDPL. Cannot be guaranteed in terms of performance, and must there be established by the operator.

Calibration

For Calibration use the "Multicalibrator"

Calibration Stability

For the instrumentation series Miura, the calibration is recommended to be done every 10 days.

Materials required but not supplied in the kit Calibrators and controls

Assay guidelines for Analyzer I.S.E. Miura

Assay guidelines for Analyzer I.S.E. Miura			
СКМВ			
Ckmb			
Kinetic-Substrate Start			
IU/L			
340 nm			
Not Use			
Reaction Volume	U.M.		
200	μΙ		
12	μΙ		
240	Sec.		
50	μΙ		
180	Sec.		
288	Sec.		
	CKMB Ckmb Kinetic-Substrate S IU/L 340 nm Not Use Reaction Volume 200 12 240 50 180		

Temperature Conversion:

Following factors can be used for conversion of IU/I from one temperature to another:

Assay	Desired temperature		
Temperature	25 ℃	30 °C	37°C
25 <i>°</i> C	1.00	1.53	2.38
30 ℃	0.65	1.00	1.56
37℃	0.42	0.64	1.00

Note: Since temperature conversion factors are given only as an approximate conversion, it is suggested that values be reported at the temperature of measurement

Normal Range

	at 25 <i>°</i> C	at 30 <i>°</i> C	at 37℃
CK-MB	<10 IU/I	<15 IU/I	<24 IU/I

Note: Expected range varies from population to population and each laboratory should establish its own normal range.

Limitations

- Avoid using haemolysed serum since red blood cells may release enzymes and intermediates such as ATP and Glucose-6phosphate dehydrogenase which may interfere in the assay.
- This procedure may over estimate CK-MB values if CK-BB activity in the serum is high. CK-BB activity is usually absent in sera from normal individual and patients with myocardial infarction. The presence of a macro form of CK-BB in the specimen should be suspected if the CK-MB activity measure by this procedure represents more than 20% of the total CK activity.
- The working solution is considered unsatisfactory and should not be used if the absorbance exceeds 0.700 at 340 nm against distilled water.
- If the CK-MB activity exceeds 2000 IU/L; then dilute the specimen suitably with normal saline and repeat the assay. In such case, the results obtained should be multiplied with the dilution factor to obtain correct CK-MB activity. The dilution of a sample that initially exceeds linearity often results in a higher than expected value; therefore instead of dilution it is recommended that a smaller sample volume be used.

Quality Control

To ensure adequate quality control measures, it is recommended that each batch should include a normal and an abnormal commercial reference control serum. It should be realized that the use of quality control material checks both instrument and reagent functions together. Factors which might affect the performance of this test include proper instrument function, temperature control, cleanliness of glassware, Wavelength setting, Expiration date of reagents and accuracy of prob aspiration.

Accuracy-Recovery

N= 96, R = 0.998, Y = 1.03x - 2.85

Interference

Triglycerides below 600 mg/dl does not interfere in the reaction. Haemoglobin interferes at concentrations above 50.0 g/L. Ascorbic Acid influences the reaction at concentrations over 64 mg/dl.

Within-run					
Range	U.M	Mean	S.D.	C.V.(%)	No. run
Low	IU/L	33.7	0.52	2.3	20
High	IU/L	125	2.15	1.8	20
Between run					
Range	U.M	Mean	S.D.	C.V.(%)	No. run
Low	IU/L	40	0.75	3.9	20
High	IU/L	135.5	2.20	1.8	20

Sensitivity

At 340 nm, the activity of CK-MB of 3 IU/L can estimate.

References

- Neumeier D,etal., Activity kinetics and Diagnostic Significance in Myocardial infarction. Clinica Chimica Acta 73; 445-451, 1976.
- 2. Gerhardt W, et al., Creatine kinase B-sub unit Activity in human serum. Clinica Chimica Acta 78:29-41, 1977.
- 3. Melattini F, et al., Clin. Chem. 24/3: 498-501, 1978.
- Wicks R, et al., Immunochemical Determination of CK-MB Isoenzyme in human serum, II. Enzyme approach Clin. Chem. 28/1; 54-58, 1982.
- 5. Tietz N. W., Fundamentals of Clin. Chem. (III) 377, 383 (1981).
- 6. Faulkner, Willard R., Selcted Methods of Clin. Chem. (9) 185 (1982).
- 7. In-house test data,

Symbols

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